# Experienced User Protocol

All spins at ;12,000 3 g, except as noted.

## 1 Harvest & lyse bacteria

- Pellet cells from 1–5 ml overnight culture *1 minute* (1 ml from TB or 2xYT; 1–5 ml from LB medium). Discard supernatant.
- $\bullet~$  Resuspend cells in 200  $\mu l$  Resuspension Solution. Pipette up and down or vortex.
- Add 200 µl of Lysis Solution. Invert gently to mix. Do not vortex. Allow to clear for :5 minutes
- \* Prior to first time use, be sure to add the RNase A to the Resuspension Solution.

## 2 Prepare cleared lysate

- Add 350 µl of Neutralization Solution (S3).
  Invert 4–6 times to mix.
- Pellet debris 10 minutes at max speed.

## 3 Prepare binding column

- Add 500 ml Column Preparation Solution to binding column in a collection tube.
- Spin at  $\geq$ 12,000 x q, 1 minute. Discard flow-through.

## 4 Bind plasmid DNA to column

- Transfer cleared lysate into binding column.
- Spin 30",1 minute. Discard flow-through.

#### 5 Wash to remove contaminants

- Optional (EndA<sup>+</sup> strains only): Add 500 μl Optional Wash Solution to column. Spin 30", 1 minute. Discard flow-through.
- Add 750 µl Wash Solution to column. *Spin 30", 1 minute.* Discard flow-through.
- Spin 1 minute to dry column.
- \* Prior to first time use, be sure to add ethanol to the concentrated Wash Solution.

### 6 Elute purified plasmid DNA

- Transfer column to new collection tube.
- Add 100 µl Elution Solution. Spin 1 minute.
- \* If a more concentrated plasmid DNA prep is required, reduce the elution volume to a minimum of 50  $\mu$ l.

### **Bacterial culture**



**Pure Plasmid DNA**